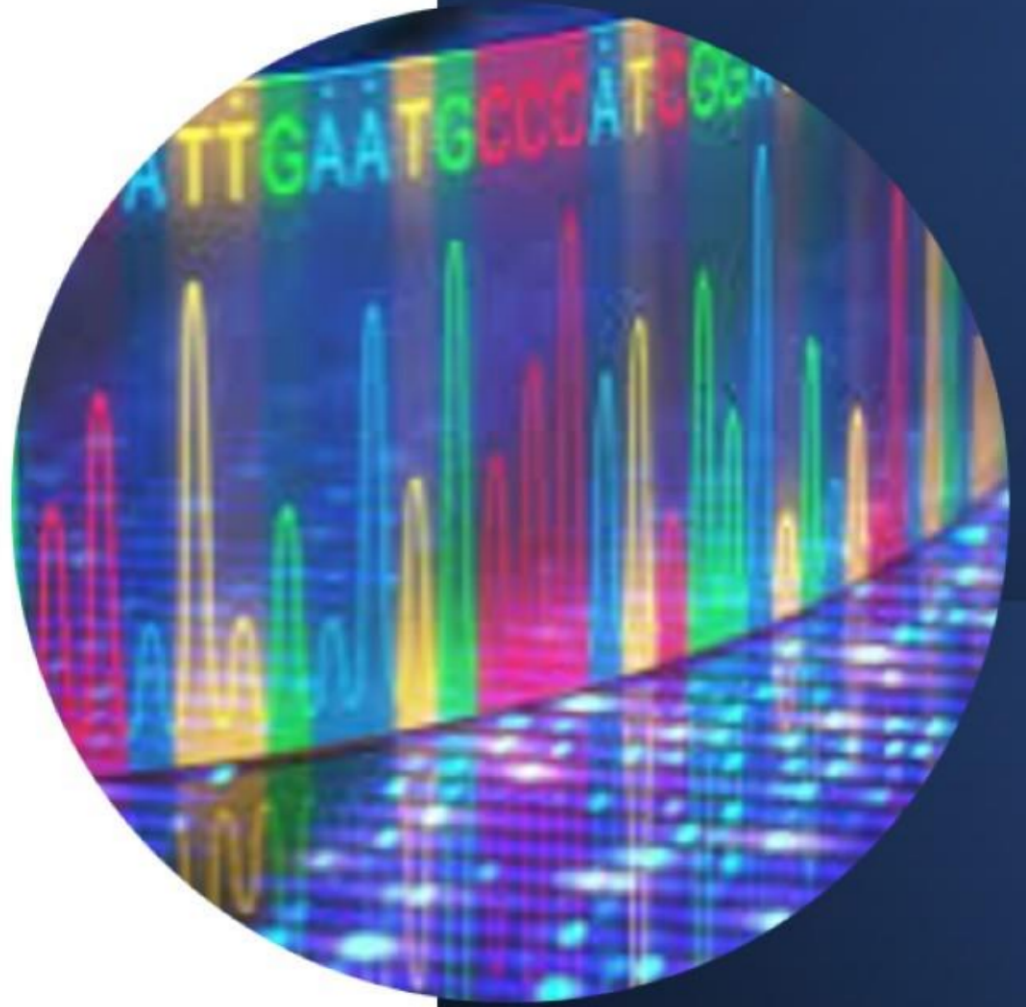


DNA sequencing

Lec 3 / Bioinformatics

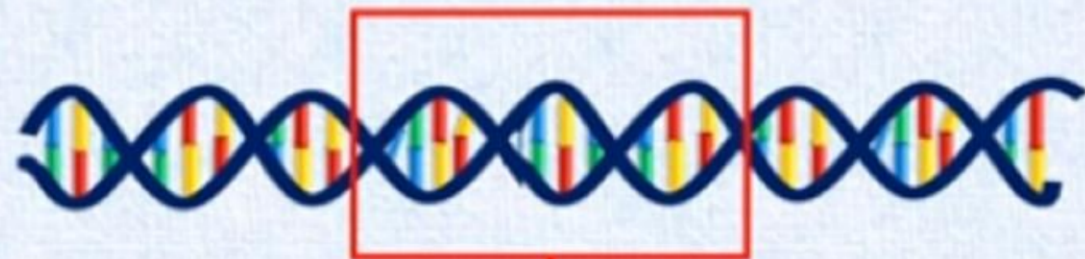
By

Dr Delveen R. Ibrahim



DNA sequencing

The technique by which the **precise order of nucleotides** in a DNA segment can be determined.



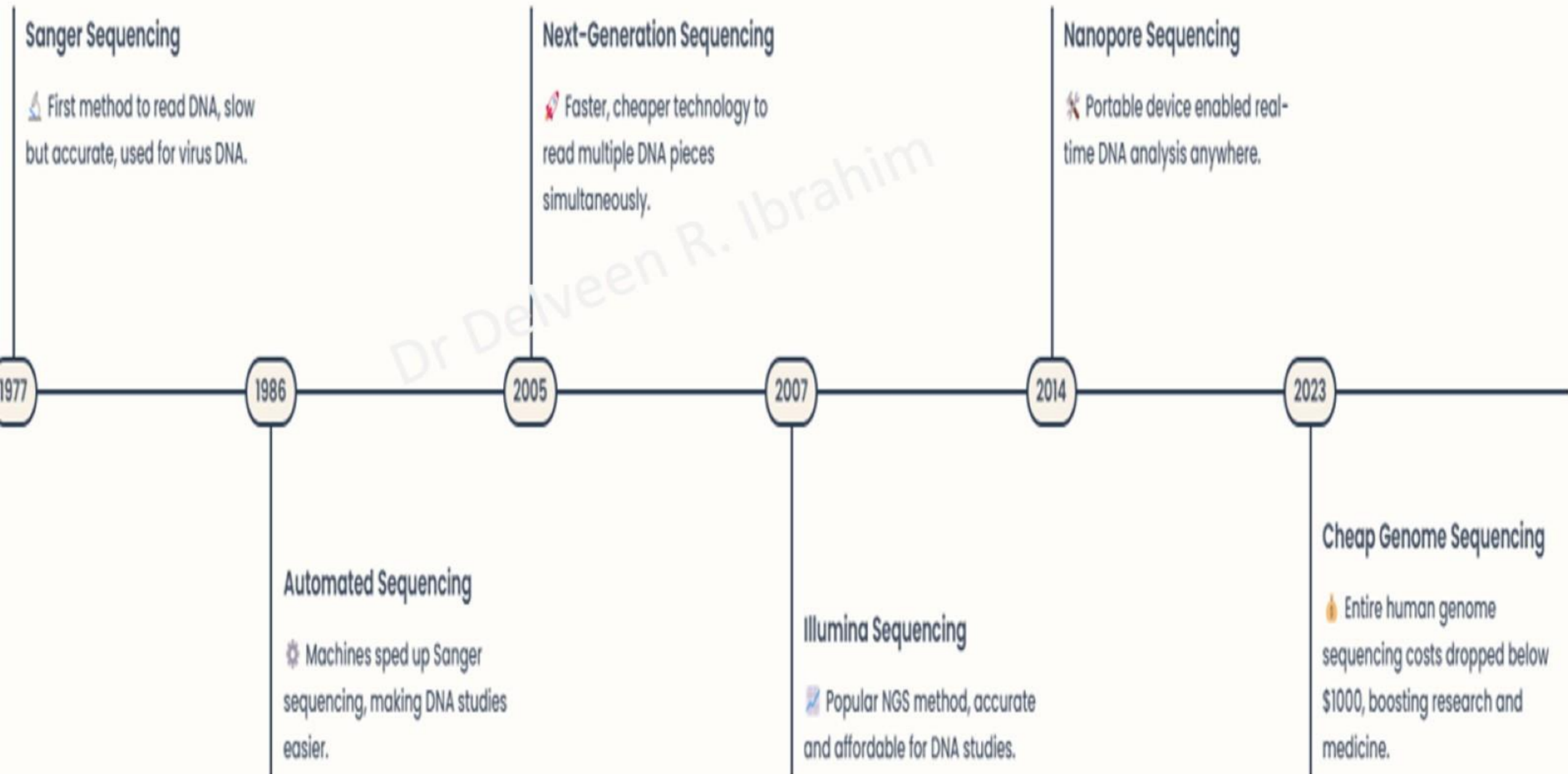
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5'ATGCACTTGATC 3'
3'TACGTGAACTAG 5'

DNA sequencing:

- Simply, Sequencing DNA means determining the order of the four chemical building blocks called "bases", that make up the DNA molecule using laboratory technique .
- In the DNA double helix, the four chemical nucleotide or bases always bond with the same partner to form "base pairs."
- These bases are Adenine, Cytosine, Thymine, Guanine.
- Adenine (A) always pairs with thymine (T); cytosine (C) always pairs with guanine (G).
- So, the role of DNA sequencing is to understand and interpret the genetics code to all biological life on earth as well as to understand and treat genetic diseases.

DNA Sequencing Developments



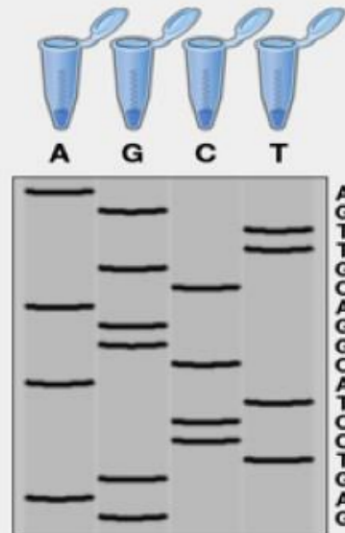
DNA Sequencing methods: there are many methods which are using different mechanisms for sequencing

DNA sequencing by synthesis

Polymerase-based DNA sequencing

Sanger DNA sequencing

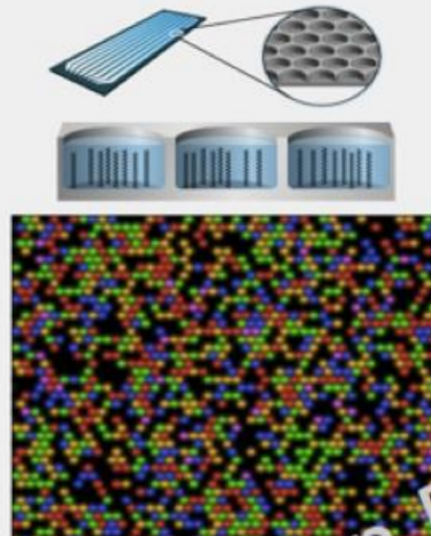
Sequence 500 - 700 DNA bases per reaction
16 reactions per gel



Sequence 10,000 DNA bases per gel

Massively parallel DNA sequencing

Sequence 100 - 5,000 DNA bases per reaction
10 thousand to 10 billion reactions per slide

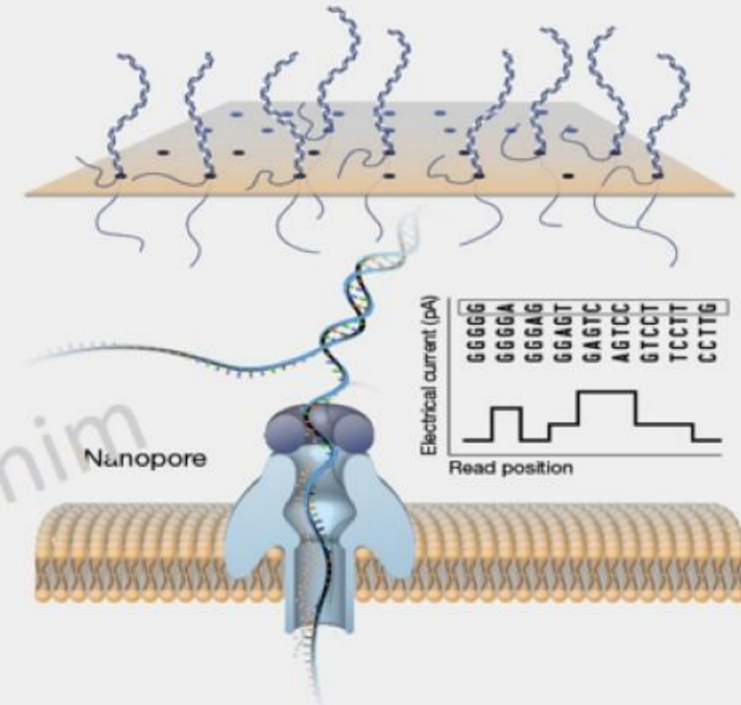


Sequence 2 trillion DNA bases per slide

Single molecule DNA sequencing

Nanopore DNA sequencing

Sequence 10 thousand to 4 million DNA bases per pore
40,000 - 250,000 pores per device



Sequence upwards of 200 billion DNA bases per device

1. Basic methods:

Category	Method	Key Features	Advantages	Limitations
Basic Methods	Maxam-Gilbert Sequencing (1977)	Uses chemicals to cut DNA at specific bases.	High accuracy for short DNA	Labor-intensive, toxic chemicals
	Sanger Sequencing (1977)	Uses special bases to stop DNA copying at different points.	Reliable and accurate	Slow, expensive for large genomes

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2. Advanced methods:

Category	Method	Key Features	Advantages	Limitations
Advanced Methods	Automated Sanger (1986)	Uses fluorescent labels and machines for faster reading.	Faster than manual Sanger	Still costly for large-scale sequencing
	Pyrosequencing (1996)	Detects light signals as DNA bases are added.	Faster than Sanger	Works best for short sequences

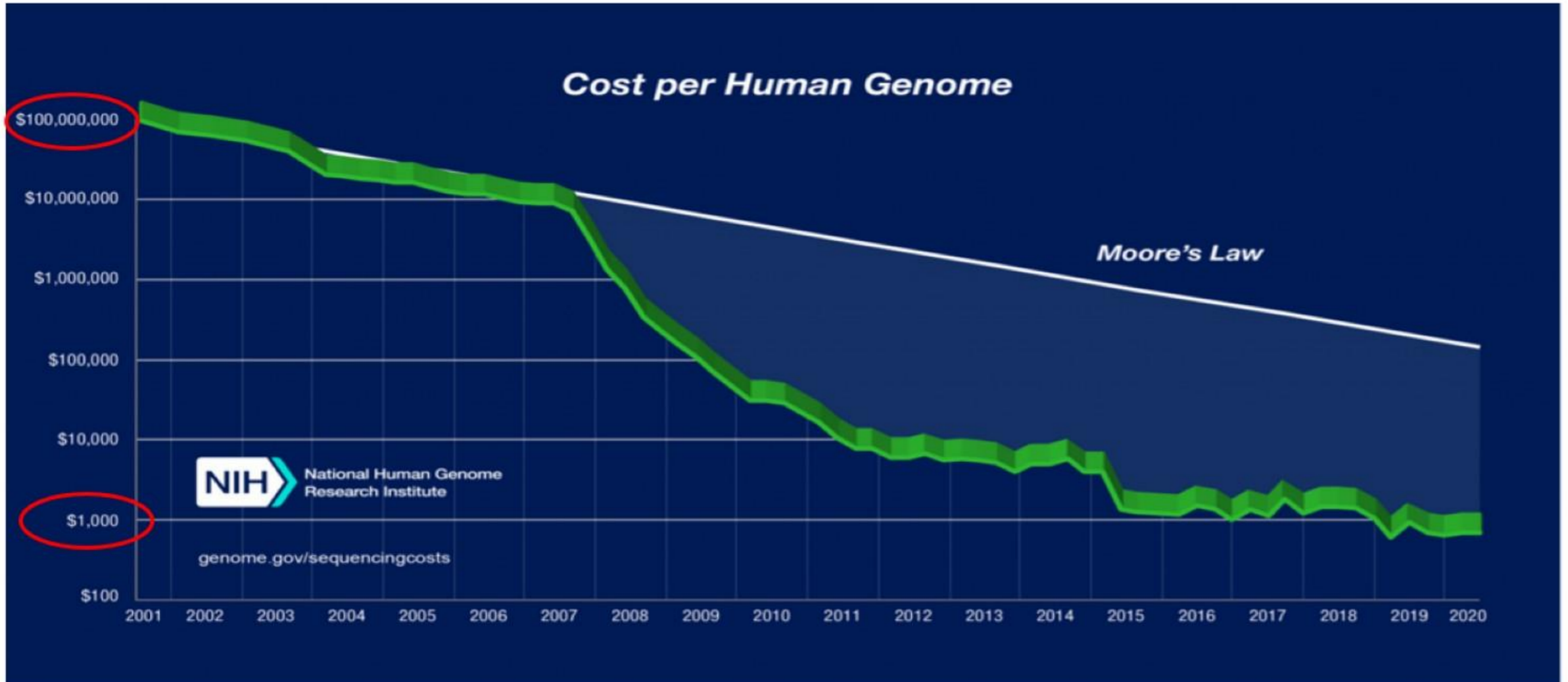
3. Next Generation Sequencing:

Category	Method	Key Features	Advantages	Limitations
Next-Generation Sequencing (NGS)	Illumina Sequencing (2007)	Uses fluorescent signals to read short DNA fragments.	High accuracy, widely used	Short read lengths
	Ion Torrent Sequencing (2011)	Detects pH changes when bases are added.	Faster and cheaper than Illumina	Less accurate for long reads

Category	Method	Key Features	Advantages	Limitations
Third-Generation Sequencing	PacBio (SMRT Sequencing, 2010)	Reads long DNA fragments in real time.	Long reads, real-time sequencing	High error rate, expensive
	Nanopore Sequencing (2014)	Uses tiny pores to read DNA as it passes through.	Portable, real-time, long reads	Higher error rates

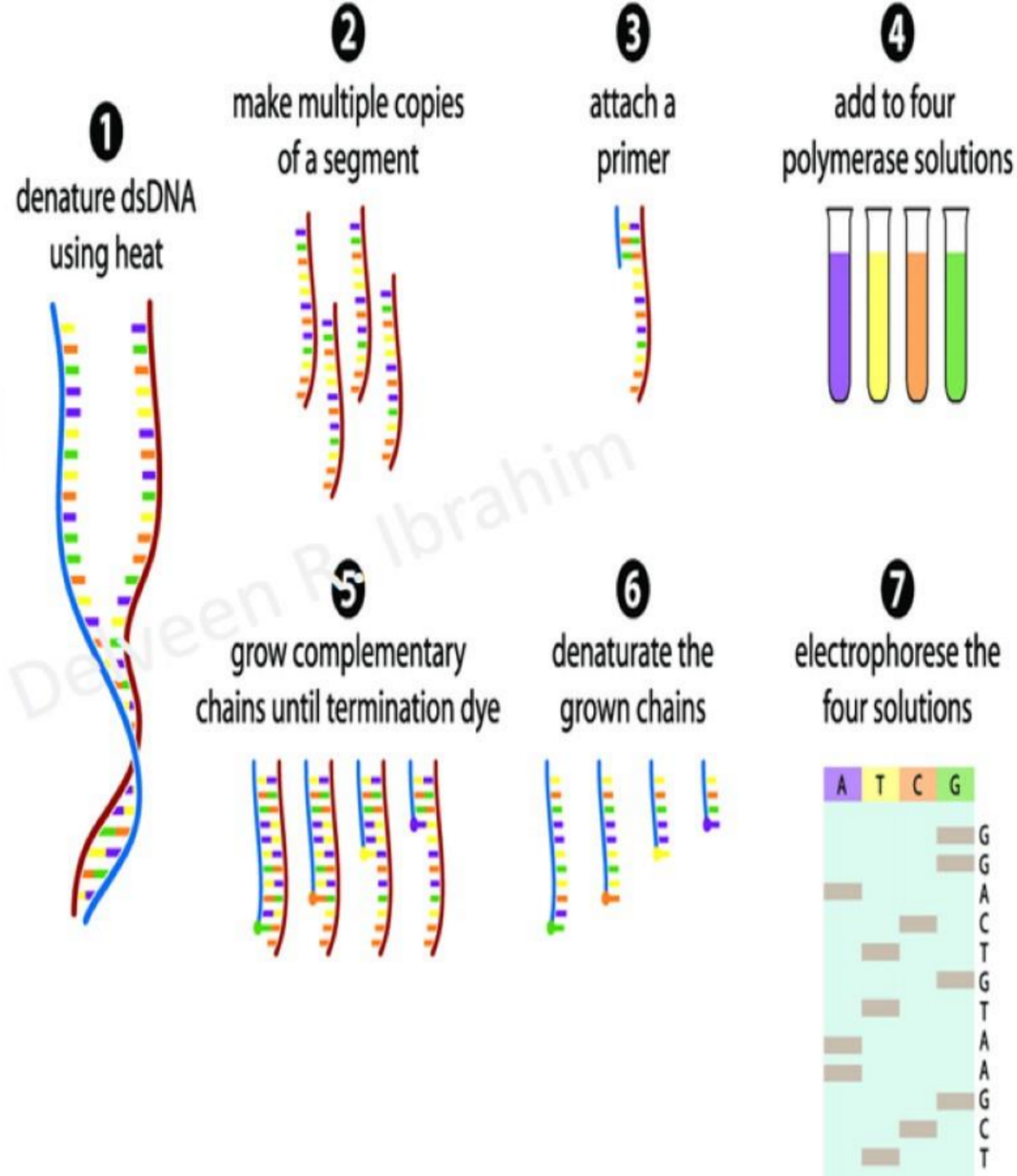
Note: Long reads and **short reads** refer to the length of DNA sequences that a sequencing method can read at a time

Due to the advanced methods used in DNA sequencing the cost also has been reduced tremendously which make it more affordable, see the figure below

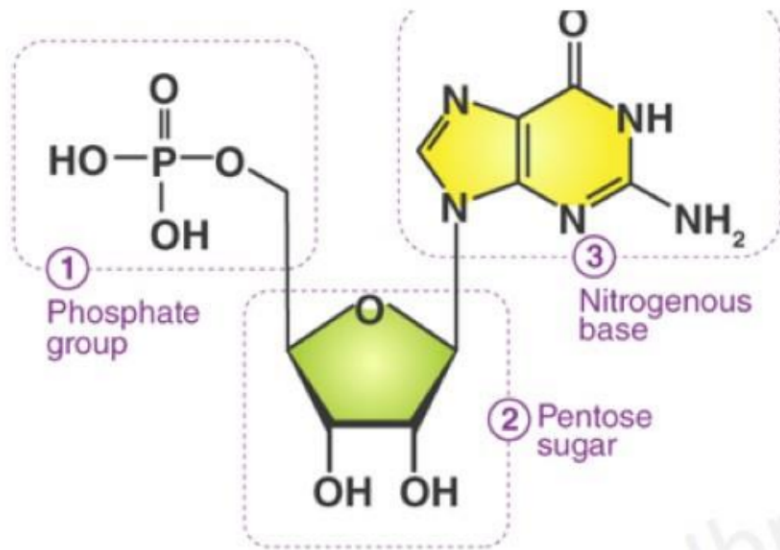


Sanger (chain termination method)

- This method includes many steps starting with DNA extraction, purification, PCR, fragment separation, detection and data analysis. See the provided video in Moodle.
- But mainly this process depends on **DNA Polymerase** and **Dideoxynucleotides (ddNTPs)**: DNA polymerase, the enzyme responsible for synthesizing new DNA strands, is then introduced along with a mixture of standard deoxynucleotides (dNTPs) and small amounts of modified nucleotides called **dideoxynucleotides (ddNTPs)**. These ddNTPs lack a 3'-OH group, which is necessary for DNA strand elongation. So, after involving one of the ddNTPs, termination occurs.

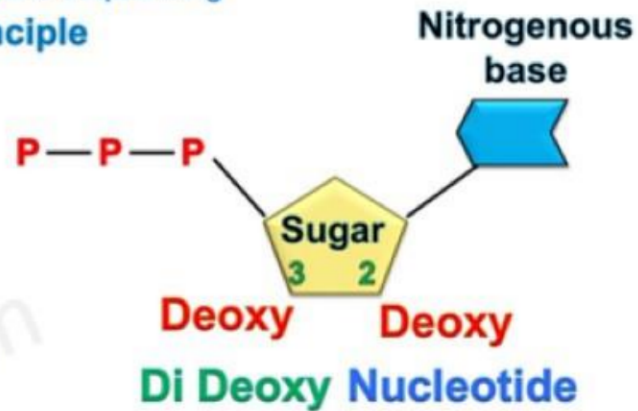


Nucleotide







Di Deoxy Nucleotide

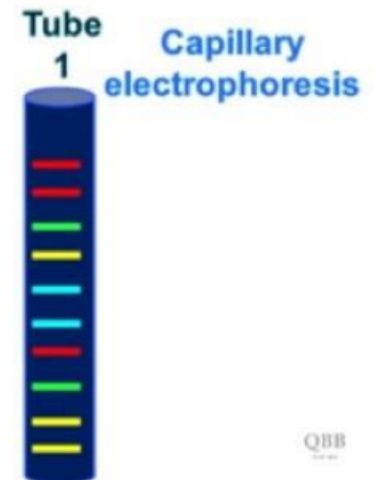
Sanger's sequencing
Principle



Fragment separation

ddATP - 
ddTTP - 
ddGTP - 
ddCTP - 

Tube 1





Sanger Sequencing Original Method

Labour Intensive



4 Days



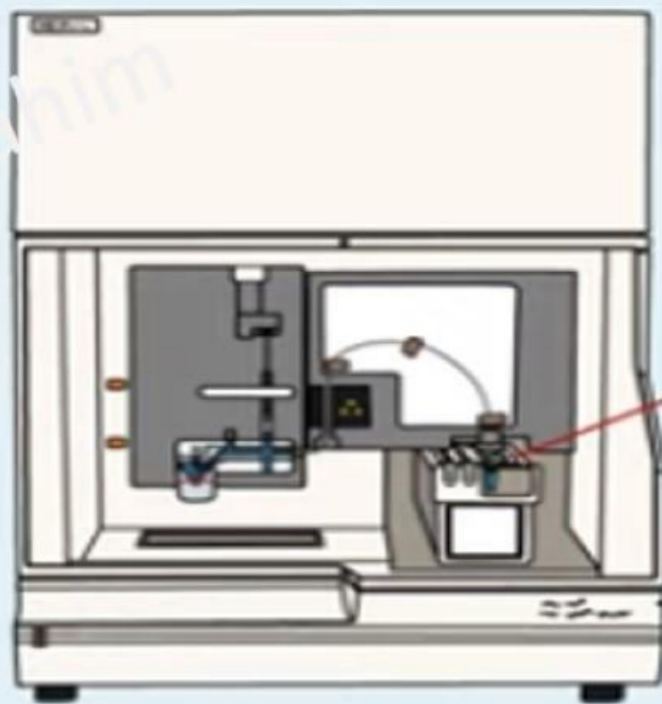
↓
 200 Nucleotides

↓
 Few Samples

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Modern Sanger sequencing

Sanger Sequencing By Capillary Electrophoresis




Read Length:
600 bp

Automated
Sample Loading
≤ 96 Samples

loading. Up to 96 samples could be loaded in a
plate on the system and left to run on its own.

Sanger Sequencing

Why Use Sanger Sequencing Today?

	 Sanger Sequencing	NGS
Accuracy	99.9 %	99 - 99.9 %
Cost Effectiveness	< 20 Samples	> 20 Samples
Speed < 20 Samples	Fast	Slow
Speed > 20 Samples	Slow	Fast
Sensitivity	15 - 20 %	1 %
Sample Coverage	1 Read/Sample (300 - 850 bp)	Billions of Reads/Sample (Up to 16 Tb)



Retrieving nucleotide and protein sequence using NCBI

Practical / Lab 3

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Finding Nucleotide sequence by using **accession number**, or **key words** such as the gene name and organism by searching the Nucleotide section in NCBI

On left upper side select **Nucleotide**. Type accession number , name of gene or gene symbol and organism name

The screenshot displays the NCBI (National Center for Biotechnology Information) homepage. At the top, the NIH logo and the text "National Library of Medicine National Center for Biotechnology Information" are visible. A search bar is located at the top right, with a "Search" button. Below the search bar, a dropdown menu is open, showing a list of search categories. The "Nucleotide" option is highlighted in blue. The main content area features a large "NCBI" heading and a brief description of the center's mission. Below this, there are three columns of links: "Submit" (with links for Manuscripts, Preprints, and Sequences), "Download" (with a link for "Transfer NCBI data to your computer"), and "Learn" (with links for "Find help documents", "attend a class", and "watch a tutorial"). At the bottom, there are three more columns: "Develop" (with a link for "Use NCBI APIs and code libraries to build applications"), "Analyze" (with a link for "Identify an NCBI tool for your data analysis task"), and "Research" (with a link for "Explore NCBI research and collaborative projects"). On the left side, a vertical navigation menu lists various resources, including "NCBI Home", "Resource List (A-Z)", "All Resources", "Chemicals & Bioassays", "Data & Software", "DNA & RNA", "Domains & Structures", "Genes & Expression", "Genetics & Medicine", "Genomes & Maps", "Homology", "Literature", "Proteins", "Sequence Analysis", "Taxonomy", and "Training & Tutorials". On the right side, there are sections for "Popular Resources" (including PubMed, Bookshelf, PubMed Central, BLAST, Nucleotide, Genome, SNP, Gene, Protein, and PubChem) and "NCBI News & Blog" (including a link for "RefSeq Release 216" dated 17 Jan 2023).

NIH National Library of Medicine
National Center for Biotechnology Information

Search

NCBI

Submit
Download
Learn

Develop
Analyze
Research

Popular Resources

NCBI News & Blog

Type *gyrA* (gyrase subunit A) for example, you will get the window below. Click on the first option , you will get the GenBank and you can do copy the complete gene or the CDS just like before

Nucleotide [Create alert](#) [Advanced](#)

Species [Summary](#) [20 per page](#) [Sort by Default order](#) [Send](#)

Animals (54)
Plants (88)
Fungi (32)
Protists (70)
Bacteria (1,329,512)
Archaea (3,035)
Viruses (158)
Customize ...

Molecule types
genomic DNA/RNA (1,332,920)
mRNA (34)
rRNA (2)
Customize ...

Source databases
INSDC (GenBank) (870,980)
RefSeq (274,880)
Customize ...

Sequence Type
Nucleotide (1,333,102)
EST (14)
GSS (8)
Genetic compartments

See [GYRA DNA GYRASE A](#) in the Gene database
[gyrA](#) reference sequences [Transcript \(1\)](#) [Protein \(1\)](#)

Items: 1 to 20 of 1333124

☐ [Mycobacterium tuberculosis strain UKR100 GyrA \(gyrA\) gene, complete cds](#)

1. 2,517 bp linear DNA
Accession: MG995190.1 GI: 1373737658
[Protein](#) [Taxonomy](#)
[GenBank](#) [FASTA](#) [Graphics](#) [PopSet](#)

☐ [Mycobacterium tuberculosis strain UKR99 GyrA \(gyrA\) gene, complete cds](#)

2. 2,517 bp linear DNA
Accession: MG995189.1 GI: 1373737656
[Protein](#) [Taxonomy](#)
[GenBank](#) [FASTA](#) [Graphics](#) [PopSet](#)

☐ [Mycobacterium tuberculosis strain UKR98 GyrA \(gyrA\) gene, complete cds](#)

3. 2,517 bp linear DNA
Accession: MG995188.1 GI: 1373737654

GenBank

Mycobacterium tuberculosis strain UKR100 GyrA (gyrA) gene, complete

GenBank: MG995190.1

[FASTA](#) [Graphics](#) [PopSet](#)

Go to:

LOCUS MG995190 2517 bp DNA linear BCT 03-APR-2018
DEFINITION Mycobacterium tuberculosis strain UKR100 GyrA (gyrA) gene, complete cds.
ACCESSION MG995190
VERSION MG995190.1
KEYWORDS .
SOURCE Mycobacterium tuberculosis (Mycobacterium tuberculosis variant tuberculosis)
ORGANISM [Mycobacterium tuberculosis](#)
Bacteria; Actinomycetota; Corynebacteriales; Mycobacteriaceae; Mycobacterium; Mycobacterium tuberculosis complex.
REFERENCE 1 (bases 1 to 2517)
AUTHORS Daum, L.T. and Rodriguez, J.D.
TITLE Next-Generation Sequencing for Characterizing Drug Resistance-Confering Mycobacterium tuberculosis Genes from Clinical Isolates in the Ukraine
JOURNAL J. Clin. Microbiol. (2018) In press

What are the prefix means in accession numbers?

- . **NM_**: Refers to mRNA sequences (messenger RNA).
- . **NP_**: Refers to protein sequences (translated products of mRNA).
- . **NG_**: Refers to incomplete genomic regions (usually gene-specific loci).
- . **NC** stands for **Non-redundant Curated** sequence



Protein



CCR5



Search

NCBI Home

Resource List (A-Z)

All Resources

Chemicals & Bioassays

Data & Software

DNA & RNA

Domains & Structures

Genes & Expression

Genetics & Medicine

Genomes & Maps

Homology

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Sequence Analysis

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Nucleotide

Genome

SNP

Gene

Protein

PubChem

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Go to Settings to activate Windows.

NCBI News & Blog

New! Introducing the Multiple

[Custom range...](#)[Molecular weight](#)[Custom range...](#)[Release date](#)[Custom range...](#)[Revision date](#)[Custom range...](#)[Clear all](#)[Show additional filters](#)[RefSeq products](#)[Orthologs](#)[Genome Data Viewer](#)

New - Visualize gene across multiple species

RefSeq SequencesDatabase: [Select](#)[Find items](#)

Search details

CCR5[All Fields]

[Search](#)[See more...](#)**Items: 1 to 20 of 15792**<< First < Prev Page **1** of 790 Next > Last >>☐ [ccr5 \[Homo sapiens\]](#)

1. 352 aa protein

Accession: AAB57793.1 GI: 2104520

[Nucleotide](#) [Taxonomy](#)[GenPept](#) [Identical Proteins](#) [FASTA](#) [Graphics](#)☐ [CCR5, partial \[Homo sapiens\]](#)

2. 166 aa protein

Accession: ADC94815.1 GI: 289466095

[Nucleotide](#) [Taxonomy](#)[GenPept](#) [Identical Proteins](#) [FASTA](#) [Graphics](#)☐ [CCR5, partial \[Rattus rattus\]](#)

Recent activity

[Turn Off](#) [Clear](#)

CCR5 (15792)

Protein

Homo sapiens BRCA1 DNA repair associated (BRCA1), transcript varia Nucleotide

Homo sapiens BRCA1 DNA repair associated (BRCA1), RefSeqGene Nucleotide

breast cancer type 1 susceptibility protein isoform 147 [Homo sapiens] Protein

Homo sapiens chromosome 17, GRCh38.p14 Primary Assembly Nucleotide



Protein

Protein ▼

Search

Advanced

Help

GenPept ▼

Send to: ▼

Change region shown ▼

Customize view ▼

ccr5 [Homo sapiens]

GenBank: AAB57793.1

[Identical Proteins](#) [FASTA](#) [Graphics](#)

Go to: ▼

LOCUS	AAB57793	352 aa	linear	PRI 26-JUL-2016
DEFINITION	ccr5 [Homo sapiens].			
ACCESSION	AAB57793			
VERSION	AAB57793.1			
DBSOURCE	locus HSU95626 accession U95626.1			
KEYWORDS	.			
SOURCE	Homo sapiens (human)			
ORGANISM	Homo sapiens Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini;			

Analyze this sequence ▼

Run BLAST

Identify Conserved Domains

Highlight Sequence Features

Find in this Sequence

Protein 3D Structure ▼

Activate Windows
Go to Settings to activate Windows.

NMR solution structure of monomeric CCL5 in complex

Tasks:

You will be given different tasks in the lecture